

FLUOROQUINOLONE RESISTANCE IN CLINICAL AVIAN PATHOGENIC ESCHERICHIA COLI ISOLATES FROM FLANDERS (BELGIUM)

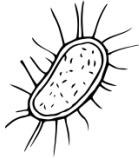
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Introduction – Objectives



Colibacillosis is one of the leading causes of **disease-related economic loss** in the **poultry sector**



Colibacillosis is caused by an infection of **avian pathogenic *Escherichia coli* (APEC)**



Fluoroquinolones, such as enrofloxacin (ENRO) are frequently used antimicrobials (via the drinking water) for the treatment of APEC infections in the poultry industry



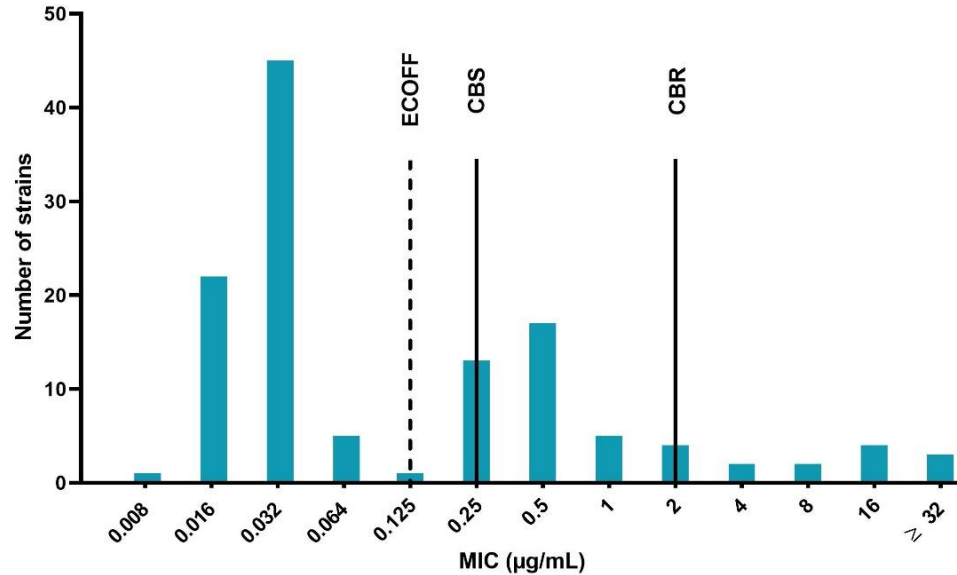
However, development and selection of **resistance** to these antimicrobial drugs is an increasing problem



The goal of this study was to assess the **prevalence and mechanisms of resistance** against ENRO in clinical APEC isolates from Flanders.

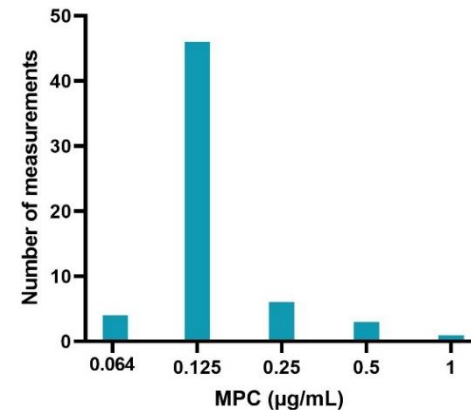
- Determination of minimum inhibitory concentration (**MIC**) distribution of 125 clinical APEC isolates (gradient strip test)
- Determination of mutant prevention concentration (**MPC**) values of a sample of sensitive strains, i.e. MIC values of 0.016 and 0.032 µg/mL (agar dilution technique)
- Non-wild type (NWT) strains were further screened for the presence of **QRDR** (quinolone resistance determining regions) mutations (*gyrA*, *ParC*, *ParE*) and **PMQR** (plasmid mediated quinolone resistance) genes (*qnrA*, *qnrB*, *qnrS*, *oqxAB*, *qepA*) using PCR, gel electrophoresis and gene sequencing (Sanger)

Results: MIC and MPC



← **Figure (left).** Enrofloxacin MIC distribution of the 125 avian pathogenic *Escherichia coli* (APEC) strains isolated from broiler farms in Flanders with colibacillosis outbreaks. The dotted line represents the epidemiological cut-off (ECOFF), which is 0.125 µg/mL. Strains with MIC values ≤ ECOFF are labelled wild type (WT, 60% of the strains) and strains with higher MIC values than the ECOFF as non-wild type (NWT, 40% of the strains). The full lines marked CBS and CBR indicate the clinical breakpoint for susceptibility (0.25 µg/mL, CBS) and resistance (2 µg/mL, CBR), respectively. Strains with MIC in between CBS and CBR are designated intermediate (21% of the strains). Sixty-nine percent of the strains were susceptible and 10% resistant.

→ **Figure (right)**. Enrofloxacin MPC distribution of 20 sensitive avian pathogenic *Escherichia coli* (APEC) strains obtained via agar dilution experiments, performed in triplicate (60 measurements in total).



Results: QRDR and PMQR

MIC (µg/mL)	Number of isolates	QRDR mutations			PMQR
		<i>gyrA</i>	<i>parC</i>	<i>parE</i>	
0.25	1				<i>qnrS</i>
	12	S83L			
0.5	17	S83L			
1	2				<i>qnrS</i>
	2	S83L			
	1	S83L	S80R		
2	1				<i>qnrS</i>
	2	S83L			<i>qnrB</i>
	1	S83L	S80R		
4	2	S83L			<i>qnrS</i>
8	1	S83L			<i>qnrS</i>
	1	S83L/D87N	S80R		
16	4	S83L/D87N	S80I		
≥ 32	1	S83L/D87N	S80R	S458A	
	2	S83L/D87N	S80I	S458A	

← **Table.** Overview of the fluoroquinolone resistance mechanisms (QRDR and PMQR) of the different NWT strains (strains with MIC values above the ECOFF). Of all the NWT strains (total 50) :

- 92% carried one or more mutations in *gyrA*.
- *ParC* mutations occurred in 22% of the cases
- *ParE* mutations were rare (6% or 3 strains)
- 18% possessed PMQR associated genes. Most detected was *qnrS* (14%), followed by *qnrB* (4%). The other investigated genes were not detected.

Conclusion

- **MIC results:** A large number of strains (40%) were considered NWT. In contrast to the relatively high number of NWT strains, only a minority of this group was designated as resistant (R), indicating the presence of a considerable intermediate portion of strains (21%).
- **MPC results:** MPC values of the tested isolates were fairly stable across the different. In 76.67% of the cases, the MPC was 0.125 µg/mL. Approximately 17% of the measurements had higher MPC values. Two strains exhibited a MPC/MIC ratio of 32 (MPC 0.5 and 1 µg/mL, respectively), indicating a large mutant selection window (MSW). A wide MSW increases the risk of selective enrichment of resistant subpopulations in a bacterial culture.
- **QRDR/PMQR results:** Chromosomal mutations in DNA gyrase and topoisomerase IV are the main source of fluoroquinolone resistance in *E. coli*, deemphasising the role of PMQR mechanisms.

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